

## A coarse-grained simulator: CafeMol

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## CafeMol (www.cafemol.org)



- CafeMol 2.1 (2013/7)
   source & manual released
- Features are;
  - Various CG models protein/DNA/RNA multiple basin model accurate CG model
  - Simulating protein-at-work "switching"
- Under development
   lipid



## **Overview of CafeMol**



- General-purpose coarse-grained (CG) biomolecular modeling and simulation software
  - Protein: 1 bead / 1 amino acid
  - Nucleic acid:
    - 3 beads (sugar, base, phosphate) / nucleotide
  - Lipid: ~3 beads / lipid
- Written by FORTRAN90 with MPI and Open MP
- Large-scale simulation
  - ~"millisecond" event by K-computer
- Version 1.0 is released (only protein) (2010/12/27)
  - Version 2.0 (protein, DNA, RNA) (2012/5/31)

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## **Models and energy functions**





1 beads / 1 amino acid

- A. Off-lattice Go model
- B. Atomic interaction based CG model
- C. Multiple basin model
- D. DNA/RNA model
- E. Elastic network model
- F. Electrostatic and hydrophobic interactions
- G. Explicit and implicit ligands

## **Off-lattice Go model**



C. Clementi, H. Nymeyer, and J.N. Onuchic, J. Mol. Biol. (2000)

Based on the energy landscape theory Structure based

$$\begin{aligned} V_{protein} &= V_{local} + V_{go} + V_{ex} \\ V_{local} &= K_b \sum_{i} \left( r_{i,i+1} - r_{0i,i+1} \right)^2 + K_{\theta} \sum_{i} \left( \theta_i - \theta_{0i} \right)^2 \\ &+ K_{\phi}^1 \sum_{i} \left( 1 - \cos(\phi_i - \phi_{0i}) \right) + K_{\phi}^3 \sum_{i} \left( 1 - \cos 3(\phi_i - \phi_{0i}) \right) \\ V_{go} &= \varepsilon_{go} \sum_{i,j}^{native} \left[ 5 \left( \frac{r_{0ij}}{r_{ij}} \right)^{12} - 6 \left( \frac{r_{0ij}}{r_{ij}} \right)^{10} \right] \\ V_{ex} &= \varepsilon_{ex} \sum_{i,j}^{nonnative} \left( \frac{\sigma}{r_{ij}} \right)^{12} \end{aligned}$$

## Atomic interaction based CG (AICG) model





Wenfei Li

1) Contact energy  $\varepsilon_{ij}$  from pairwise all-atom (AA) energy  $E^{IJ}(R_{IJ}) = \sum_{i \in I} \sum_{j \in J} u_{AA}(r_{ij}) \qquad u_{AA}(r) = V(r) + \Delta G^{GB}_{pol}(r) + \Delta G^{SA}(r)$ 

2) Coefficients fitted by AA-derived fluctuation (23 proteins)

param	K <sub>b</sub>	K <sub>a</sub> G	k <sub>a</sub> H	k <sub>a</sub> E	k <sub>a</sub> <sup>T</sup>	k <sub>a</sub> C	$\varepsilon_{\phi}^{G}$	$\varepsilon_{\phi}^{\ H}$	$\varepsilon_{\phi}^{E}$	$\varepsilon_{\phi}^{T}$	$\varepsilon_{\phi}^{\ C}$	€ <sub>nloc</sub>
Av.	109.94	13.40	40.0 3	17.3 2	19.35	11.7 0	0.29	1.76	1.32	0.82	0.81	0.37

Test for fluctuation, structural change, & folding





### **Multiple-basin model for proteins**



K. Okazaki, N. Koga, S. Takada, J.N. Onuchic, and P.G. Wolynes, PNAS (2006)





## CG DNA model

Three interactions sites
Phosphate
Sugar
Base
Reproduce various DNA
behavior
Salt-dependent melting
Bubble formation

 Mechanical properties



T.A. Knotts IV, N.Rathore, D.C. Shwartz, and J.J. Pablo, J. Chem. Phys. (2007)

### 3SPN.1 force field



E.J. Sambrisiki, D.C. Schwartz, and J.J. de Pablo, Knotts, Biophys J. (2009)

$$\begin{aligned} V_{dna} &= V_{local} + V_{stack} + V_{bp} + V_{ex} + V_{qq} + V_{solv} \\ V_{local} &= K_{b1} \sum_{i} \left( r_{i,i+1} - r_{0i,i+1} \right)^{2} + K_{b2} \sum_{i} \left( r_{i,i+1} - r_{0i,i+1} \right)^{4} \\ &+ K_{\theta} \sum_{i} \left( \theta_{i} - \theta_{0i} \right)^{2} + K_{\phi} \sum_{i} \left( 1 - \cos(\phi_{i} - \phi_{0i}) \right) \\ V_{stack} &= 4\varepsilon_{1} \sum_{i,j}^{N_{st}} \left[ \left( \frac{\sigma_{0ij}}{r_{ij}} \right)^{12} - \left( \frac{\sigma_{0ij}}{r_{ij}} \right)^{6} \right] \\ V_{stack} &= 4\varepsilon_{1} \sum_{i,j}^{N_{st}} \left[ \left( \frac{\sigma_{0ij}}{r_{ij}} \right)^{12} - 6 \left( \frac{r_{0ij}}{r_{ij}} \right)^{10} \right] \\ V_{ex} &= 4\varepsilon_{1} \sum_{i,j}^{N_{st}} \left[ \left( \frac{\sigma_{0}}{r_{ij}} \right)^{12} - \left( \frac{\sigma_{0}}{r_{ij}} \right)^{6} \right] + \varepsilon_{1} (if \ r_{ij} < d_{cut}), \\ &= 0 (if \ r_{ij} > d_{cut}) \end{aligned}$$



# 3SPN.1 force field (electrostatic and solvation interaction)

$$V_{qq} = \sum_{i,j}^{N} \left( \frac{q_{i}q_{j}}{4\pi\varepsilon_{0}\varepsilon(T,C)r_{ij}} \right) e^{-r_{ij}/\kappa_{D}} \text{ Debye-Huckel theory}$$

$$\varepsilon(T,C) = \varepsilon(T)a(C)$$

$$\varepsilon(T) = 249.4 - 0.788T/K + 7.20 \times 10^{-4}(T/k)^{2}$$

$$a(C) = 1.000 - 0.2551C/M$$

$$+ 5.151 \times 10^{-2}(C/M)^{2} - 6.889 \times 10^{-3}(C/M)^{3}$$

$$V_{solv} = \sum_{i

$$\varepsilon_{s} = \varepsilon_{N}A_{I}$$

$$e_{N} = e_{0} (1 - [1.40418 - 0.268231N_{m}]^{-1})$$$$

 $A_{I} = 0.474876(1 + \{0.148378 + 10.9553[Na^{+}]\}^{-1})$ 

## RNA model



N. Hori and S. Takada (2012)

1 nucleotide = 3 beads



 Phosphate (phosphorus atom)
 Sugar (center of ribose ring)
 Base (pyrimidine: N1 atom) (purine: N3 atom)

# RNA model (local)



$$V_{\text{total}} = V_{\text{local}} + V_{\text{stack}} + V_{\text{basepair}} + V_{\text{nonlocal}} + V_{\text{exclude}}$$



## RNA model (nonlocal)





### **Elastic network model**





## **Electrostatic and hydrophobic interactions**

Debye-Huckel form for electrostatics

$$V_{\rm ele} = \sum_{i < j}^{N} \frac{q_i q_j}{4\pi\epsilon_0 \epsilon_k r_{ij}} e^{-r_{ij}/\kappa_D}$$

HP interactions analogous to ASA

$$V_{\rm HP} = -c_{\rm HP} \sum_{i \in \rm HP} \epsilon_{\rm HP,A(i)} S_{\rm HP}(\rho_i)$$



Count coordination number for each hydrophobic particle

## **Explicit and Implicit ligands**



Explicit ligand; as a rigid molecule



Implicit ligand; MD-MC scheme with ligand-mediated contact

$$V_{protein} \xleftarrow{k_{on}}_{k_{off}} V_{protein} + V_{imp-lig}$$

$$V_{imp-lig} = \sum_{\substack{ligand-mediated \\ contact-pairs}} - c_{lig}\varepsilon_{go} \exp\left[-\frac{(r_{ij}/r_{0ij}-1)^2}{2(\sigma/r_{0ij})^2}\right]$$

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## Simulation method

- Dynamics
  - Newtonian dynamics with Berendsen thermostat
  - Langevin dynamics
  - Multi-Particle Collision dynamics (MPC)
- Time integration
  - velocity Verlet algorithm
- Run mode
  - Constant temperature simulation
  - Simulated annealing
  - Auto-search of Tf
  - Replica exchange method
  - Potential "switching"
- Useful option
  - anchor, bridge, pulling, fix, box



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## CafeMol code





#### • Parallelization

- neighboring list, force, energy
  - →hybrid(MPI+Open MP)
- replica exchange
  - →MPI(temperature/Hamiltonian REMD)



## **Performance of MPI parallelization**

1300 base pairs DNA (7798 particles) BG/L at Riken



High parallelization efficiency

## Performance of parallelization at K-Computer



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## Native fluctuation by off-lattice Go model





## Folding simulation of src SH3 domain

N. Koga, and S. Takada, J. Mol. Biol. (2001)



# Example of input file (folding simulation of src SH3)





Sequence/structure



# Folding temperature of src SH3 (Auto-search of Tf)

Bi-section method

<<<< job\_cntl i\_run\_mode = 4 i\_simulate\_type = 1 i\_initial\_state = 1 >>>> <<<< searching\_tf tempk\_upper = 500.0 tempk\_lower = 100.0 >>>>

\*\*\*\*\*\* tf out tempk n state d state p trans tf out 300.000 995 5 \*\*\*\*\* tf\_out tempk n\_state d\_state p\_trans tf out 400.000 1000 0 \*\*\*\*\*\* tf\_out tempk n\_state d\_state p\_trans tf out 350.000 166 835 78 \*\*\*\*\*\* tf\_out tempk n\_state d\_state p\_trans tf out 325.000 953 48 19 \*\*\*\*\*\* . . . \*\*\*\*\* tf out tempk n state d state p trans tf out 341.406 638 363 98 



## Folding temperature of some proteins

Protein	Number of amino acid	Folding temperature(K)
albumin binding domain	53	380.4
src SH3 domain	56	342.9
protein G	56	338.2
$\alpha$ -spectrin SH3 domain	57	360.1
Sso7d	64	332.0
protein L	78	374.2
Im9	86	382.0
cytochrom B562	106	352.2

## "Switching" simulation







## Rotation mechanism of F<sub>1</sub>-ATPase by switching Go model



N. Koga, and S. Takada, PNAS (2006)

## Conformational change by MBP



K. Okazaki, N. Koga, S. Takada, J.N. Onuchic, and P.G. Wolynes, PNAS (2006)



<<<< unit\_and\_state  $i\_seq\_read\_style = 1$ i\_go\_native\_read\_style = 1 protein 1GGG\_2.pdb 1a 1WDN 2.pdb protein >>>> <<<< energy\_function NLOCAL(1a/1a) GO EXV NLOCAL(1b/1b) GO EXV MULTIGO\_SYSTEM(1a) 1a/1a MULTIGO\_SYSTEM(1b) 1b/1b i\_use\_atom\_protein = 0 i\_use\_atom\_dna = 0 >>>> <<<< multiple go bdemax\_mgo = 100.0 baemax mgo = 1.0 $dihemax_mgo = 0.5$ ENEGAP(1)(1) 0.0 -1.8 DELTA(1ab) 28.0 >>>>

# CafeMol

## Sliding movement of KIF1A

R. Kanada, et al PLOS Comp Biol (in press)

1 phase: multiple-basin (T, D) 2 phase: go(D) 3 phase: multiple-basin(D, phi) 4 phase: go(phi) 5 phase: go(T)

KIF1A:blue tubulin:green cargo:yellow





## **DNA duplex**

- 30 bp oligomer of DNA
- Langevin dynamics (300K)
- [Na<sup>+</sup>] = 69mM



<<<< unit\_and\_state i\_seq\_read\_style = 2 i\_go\_native\_read\_style = 3 1-2 dna sequence >>>> <<<< energy\_function DH LOCAL(1-2) L\_BDNA NLOCAL(1-2/1-2) DNA ELE  $i\_use\_atom\_protein = 0$ 3SPN.1  $i_use_atom_dna = 0$ >>>> Intra mol 1,2 <<<< electrostatic Inter mol 1-2  $cutoff_ele = 20.0$ ionic\_strength = 0.069 diele water = 78.0>>>> <<<< in box xbox = 120.0ybox = 120.0zbox = 120.0boxsigma = 4.0>>>>

## Simulation of nucleosome



Electrostatic interaction +
 Go potential

 $\epsilon_{go} \,^{pro-dna} = 0.5 \epsilon_{go} \,^{pro}$ [Na<sup>+</sup>] = 50mM

```
<<<< energy_function
NLOCAL(1-2/1-2) ELE DNA
NLOCAL(1-2/3-10) GO EXV ELE
NLOCAL(3-10/3-10) GO EXV
i_use_atom_protein = 0
i_use_atom_dna = 0
>>>>
<<<< electrostatic
cutoff_ele = 20.0
ionic_strength = 0.05
diele_water = 78.0
>>>>
```

protein-DNA Go potential H. Kenzaki, et al unpublished data

 $\epsilon_{ao}$  pro-dna:coefficient of

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